THE EFFECT OF PROPIONIC ACID ADDITION IN POULTRY DIETS ON THE INTERNAL ORGANS OF BROILER CHICKENS

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ABSTRACT

This study aims to determine the effects of adding propionic acid in the poultry diets on the internal organs of broiler chickens. The livestock used were 180 broilers with Cobb strain. This study used a completely randomized design (CRD) consisting of 3 treatments and 6 replications. The treatments in this study consisted of: P0= commercial feed without propionic acid addition; P1= commercial feed with addition of 0.5% propionic acid; and P2= commercial feed with 0.75% propionic acid. The parameters observed included live weight, spleen percentage, gizzard percentage, liver percentage, and small intestine length. The results showed that the addition of propionic acid in the broiler feed had a significant effect (P<0.05) on the live weight and had no significant effect (P>0.05) on the percentage of spleen, percentage of gizzard, percentage of liver, and small intestine length. The conclusion of this study was that the addition of propionic acid in poultry feed at a dose of 0.75% did not negatively affect the internal organs and the length of the small intestine of broiler chickens.

Key words: broiler chickens, physiological organs, propionic acid

INTRODUCTION

To produce a good broiler chicken production, efforts are needed to maintain the health of the digestive tract of the animal. Generally, farmers add Growth Promoter Antibiotic (GPA) as a feed additive. The GPA can improve digestibility in broiler chickens by maximizing absorption of nutrients in the digestive tract, reducing production of toxins from digestive tract bacteria, and reducing the occurrence of infections in the digestive tract (Julendra et al., 2010). According to Daud (2005), the use of antibiotics as additives in animal feed can leave antibiotic residues that might be toxic to consumers. These residues can create resistant microorganisms in humans and livestock, especially pathogenic bacteria such as Salmonella, Escherichia coli, and Clostridium perfringens. When these bacteria infect human or livestock bodies, the resulting diseases will be difficult to cure. Therefore, the use of a safe feed additive is very important so that there is no bad impact on consumers. The type of feed additive that is safe to use is organic acids as they leave no residue. When added to the feed, organic acids will improve the performance of the digestive organs because they increase the quality of digestive enzymes, lowering the gastrointestinal pH, and reduce the number of pathogenic bacteria in the digestive tract (Roth and Kirchgessener, 2003).

One type of organic acid that can be added to animal feed is propionic acid. This acid can improve and maintain the condition of the digestive tract of monogastric livestock. Lucke (1980) as cited in Papatsiros and Bilinis (2014) reported that organic acids commonly used in animal feed are formic, acetic, propionic, and lactic acids, all of which have specific abilities to penetrate bacterial cell walls and kill bacteria. According to Lu et al. (2003) propionic acid has the ability to inhibit fungi, increase the growth of good bacteria in the digestive tract, reduce the pH of the digestive tract so as to stimulate digestive enzyme activity, and maintain a healthy condition, and microflora balance in the digestive tract. The microflora in the digestive tract plays an important role in the productivity and health of livestock, absorption of nutrients, pathogenicity, and immunity. According to Hardy (2003), propionic acid can carry out the ionization process easily by releasing hydrogen. The increase in the number of hydrogen ions will reduce the pH of the digestive tract of monogastric livestock, so that microorganisms that cannot tolerate acidic conditions will experience slow growth or die. Khosravi et al. (2012) reported that propionic acid can be used as a substitute for GPAs in broiler feed during maintenance.

The ability of propionic acid to maintain the condition of the digestive tract might affect the activity of internal organs that play an important role in the process of absorption of feed substances. Rimba wanto et al. (2019) reported that the addition of natural acidifier-based organic acids in the form of citric acid and lactic acid in poultry feed showed the same effect on the
weight of small intestine and internal organs of broiler chickens. This is because the absorption process of food substances runs normally due to unaffected weight of chicken internal organs. There are still few studies that discuss the use of propionic acid as a feed additive in poultry and its effects on physiological organs and the digestive tract. A good digestive tract condition indicates a good condition of the body and digestive organs.

**MATERIALS AND METHODS**

This study used 180 day-old broiler chickens (DOCs) strain Cobb produced by the PT. Satwa Unggas Jaya. The DOCs were randomly assigned into 10 experimental cage plots consisting of 10 animals each. The equipments used were feeding and drinking containers, postal cages seized 120 cm x 150 cm x 65 cm, artificial heater (brooding), digital scale, cutting knife, ruler or measuring tape and chicken baskets. The feeds used were HI-Pro (PT. Charoen Pokphand Indonesia Tbk) given to the chicken in the starter-grower phase (aged 1-21 days), MRI-P (PT. Cj Cheiljedang Feed Lampung) given to the chicken at the finisher phase (aged 22 to 28 days), and propionic acid (Zetox) produced by PT. Sehat Cerah Indonesia. The feeds were given according to the treatment whereas the water was given ad libitum. The nutrient composition of the feeds used is presented in the Table 1.

**Research Methods**

This study used a completely randomized design (CRD) consisting of 3 treatments and 6 replications. The treatment in this study was the addition of propionic acid in the feed, namely: P0= commercial feed without the addition of propionic acid, P1= commercial feed + propionic acid 0.5%, P2= commercial feed + propionic acid 0.75%. Before the chickens were brought in, the cages were first cleaned and disinfected. The litter mat was prepared by using the husks. The cage equipments were cleaned with water that had been added with disinfectant. The length of ileum is measured from the bile duct junction to the diverticulum to the cecum branching (Hamsah, 2013). The length of jejunum was measured from the base of the gizzard to the bile duct junction. The length of duodenum was measured from the base of the neck and the knee joints, the abdominal cavity was opened, and the entire digestive organs was collected by holding the proventiculus and pulling out the entire digestive tracts. The internal organs (liver, gizzard, and spleen) and digestive tract were separated and cleared from fatty tissue. The length of the intestine was measured and the percentage of weight internal organs to the live weight was calculated.

**Observed Variables**

The live weight, spleen percentage, gizzard percentage, liver percentage, and small intestine length were measured using the following formula: Live weight (g), obtained from weighing the chickens before slaughter and after 6-hour fasting; Spleen percentage, obtained by dividing spleen weight (g) with live weight (g) multiplied by 100%; Gizzard percentage, obtained by dividing gizzard weight (g) with live weight (g) multiplied by 100%; Liver percentage, obtained by dividing liver weight (g) with live weight (g) multiplied by 100%; and Small intestine length (cm), including the duodenum, jejunum and ileum was determined by using a measuring tape. The length of duodenum was measured from the base of the gizzard to the bile duct junction. The length of jejunum was measured from the bile duct junction to the Meckel’s diverticulum. The length of ileum is measured from the Meckel’s diverticulum to the cecum branching (Hamsah, 2013).

**Data Analysis**

The data obtained were statistically analyzed using analysis of variance. If the treatment showed a

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**Table 1. Nutrient composition of broiler chicken feed during the study**

<table>
<thead>
<tr>
<th>Feed substances</th>
<th>Feed HI-PRO*</th>
<th>Feed MR1-P**</th>
</tr>
</thead>
<tbody>
<tr>
<td>Water content (max.) (%)</td>
<td>13.0</td>
<td>13.0</td>
</tr>
<tr>
<td>Protein (%)</td>
<td>22.0-23.0</td>
<td>21.5-23</td>
</tr>
<tr>
<td>Crude fiber (max.) (%)</td>
<td>5.0</td>
<td>4.0</td>
</tr>
<tr>
<td>Fat (min.) (%)</td>
<td>5.0</td>
<td>8.0</td>
</tr>
<tr>
<td>Ash (max.) (%)</td>
<td>7.0</td>
<td>6.5</td>
</tr>
<tr>
<td>Ca (%)</td>
<td>0.9</td>
<td>0.9-1.2</td>
</tr>
<tr>
<td>P (%)</td>
<td>0.6</td>
<td>0.7-1.0</td>
</tr>
<tr>
<td>EM Kcal/kg</td>
<td>3020-3120</td>
<td>3000-3100</td>
</tr>
</tbody>
</table>

Source: *PT. Charoen Pokphand Indonesia **PT. Cj Cheiljedang Feed Lampung

**Table 2. Average live weight of broiler chickens at the end of the study**

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Live weight (g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>P0</td>
<td>1315.00±38.50\textsuperscript{a}</td>
</tr>
<tr>
<td>P1</td>
<td>1574.20±37.20\textsuperscript{b}</td>
</tr>
<tr>
<td>P2</td>
<td>1619.30±65.60\textsuperscript{b}</td>
</tr>
</tbody>
</table>

\textsuperscript{a,b}Different superscripts within the same column indicate significant different (P<0.05), P0= Without the addition of propionic acid), P1= With the addition of propionic acid of 0.5%), P2= With the addition of propionic acid of 0.75%
significant difference, then a further test is carried out using the Duncan Multiple Range Test.

RESULTS AND DISCUSSION

Effect of Treatment on Live Weight
The effect of the addition of propionic acid on the live weight of broilers is presented in Table 2. The results of the analysis of variance showed that the addition of propionic acid had a significant effect (P<0.05) on the live weight of broilers. This is because the addition of propionic acid in the feed can increase feed consumption, thereby increased live weight at the end of the study. The feed consumption in this study was 73.94 g/chicken/day, 75.64 g/chicken/day and 75.87 g/chicken/day in P0, P1, and P2, respectively. The increase in feed consumption was caused by propionic acid effect to improve the performance of digestive enzymes, thus shortening the time to digest food substances. Gautier (2002) stated that organic acids can increase the rate of feed flow so that gastric emptying is faster and causes an increase in feed consumption during. Rasyaf (2011) reported that feed consumption is an illustration of the entry of a number of nutrient elements into the body of broiler chickens.

Further test results showed that the live weight of broiler chickens in P0 (control) was significantly different (P<0.05) from those in group P1 and P2. This is because the addition of propionic acid in the feed was able to maintain the pH of the digestive tract to remain acidic. Acidic pH conditions lead to the death of pathogenic bacteria as pathogenic bacteria cannot withstand these acidic conditions. A low pH value increased the growth of good bacteria in the digestive tract. Emma et al. (2009) reported that giving lime juice containing citric acid at the level of 0.4% and 0.8% was able to increase the number of lactic acid bacteria and reduce Salmonella sp. in the digestive tract of broiler chickens and is able to maintain an acidic pH in the digestive tract. According to Mabelebele et al. (2014), the pH of the digestive tract of broiler chickens ranges from 3.47 (gizzard) to 6.43 (small intestine). Saputra et al. (2013) reported that organic acids are able to maintain the pH of the digestive tract (cache, ventriculus and intestines), that in turn suppress the growth of pathogenic bacteria and increase lactic acid bacteria which contribute to the digestive process, so that protein utilization is good for absorption of nutrients that will be used for form muscle tissue. This is in line with the opinion of Naseri et al. (2012) that organic acids act as growth promoters capable of suppressing the growth of acid-intolerant bacteria such as Escherichia coli, Salmonella spp., and Clostridium perfringens. According to Huyghebaert (2005), reducing the population of pathogenic bacteria reduces the competition for nutrient use against the host and increases the population of lactic acid bacteria. Lactic acid bacteria help the digestive protease enzyme in the intestine, maintain intestinal health, and help absorption of nutrients (Islam et al., 2008).

Treatment P1 (addition of 0.5% propionic acid) was not significantly different (P>0.05) from P2 (addition of propionic acid 0.75%) to the percentage of live weight. This is because the addition of 0.5% propionic acid is able to maintain the pH of the digestive tract, allowing the digestive process to run well. Afscharmanesh and Porreza (2005) stated that the addition of organic acids can maintain the acidic pH of the digestive tract thereby increasing the activity of the pepsin enzyme. Pepsin enzyme functions to break down protein into amino acids and is absorbed by the body, so that the process of forming muscle tissue is good and can increase body weight gain in broiler chickens. This is in line with the research results of Radhiyani et al. (2017) which reported that the addition of 0.75% acetic acid in the feed resulted in better body weight compared to the control groups. Islam et al. (2008) stated that the administration of 0.5% citric acid showed the best final body weight results.

Effect of Treatment on Percentage of Internal Organs of Broiler Chickens
The average effect of the addition of propionic acid on the percentage of internal organs (spleen, gizzard, and liver) of broilers can be seen in Table 3. The results of the analysis of variance showed that the addition of propionic acid had no significant effect (P>0.05) on the percentage of internal organs of the broilers, indicating that propionic acid did not have any negative impact on the internal organs of broilers. This is because propionic acid’s ability to suppress the growth of

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Spleen percentage (%)</th>
<th>Gizzard percentage (%)</th>
<th>Liver percentage</th>
</tr>
</thead>
<tbody>
<tr>
<td>P0</td>
<td>0.10±0.04</td>
<td>1.67±0.39</td>
<td>2.54±0.34</td>
</tr>
<tr>
<td>P1</td>
<td>0.14±0.02</td>
<td>1.59±0.22</td>
<td>2.24±0.36</td>
</tr>
<tr>
<td>P2</td>
<td>0.15±0.03</td>
<td>1.47±0.17</td>
<td>2.42±0.26</td>
</tr>
</tbody>
</table>

P0= Without the addition of propionic acid, P1= With the addition of propionic acid of 0.5%, P2= With the addition of propionic acid of 0.75%

Table 4. Length of the small intestine

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Duodenum (cm)</th>
<th>Jejunum (cm)</th>
<th>Ileum (cm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>P0</td>
<td>28.67±2.25</td>
<td>70.50±8.36</td>
<td>56.67±6.19</td>
</tr>
<tr>
<td>P1</td>
<td>29.33±2.87</td>
<td>71.83±9.87</td>
<td>57.83±6.76</td>
</tr>
<tr>
<td>P2</td>
<td>31.50±5.39</td>
<td>77.83±7.39</td>
<td>59.17±6.43</td>
</tr>
</tbody>
</table>

P0= Without the addition of propionic acid, P1= With the addition of propionic acid of 0.5%, P2= With the addition of propionic acid of 0.75%
pathogenic or toxic microorganisms so that the performance of internal organs becomes normal and the metabolism runs well. According to Jamilah et al. (2014), the addition of organic acids is able to improve the immunity of livestock so that macrophages function properly and are able to kill antigens before they reach the bloodstream to potentially inhibit the work of internal organs. The good immunity of broiler chickens causes less protein to be used for tissue repair and antibody formation, so that protein will be deposited into the tissue for the better growth. Amrullah (2004) stated that health conditions, absorption and slow flow rates will affect the size of the digestive tract; it becomes heavier, longer, and thicker.

The average percentage of spleen weight in this study is still in the normal range (0.10%-0.15%). According to Resnawati (2010), normal spleen weight in broilers ranges from 0.08 to 0.18% of live weight, whereas according to Hermana et al. (2008) the spleen weight percentage of broiler chickens aged five weeks ranges from 0.09%-0.14% and the spleen percentage of broiler chickens aged 35 days with the provision of various acidifiers and probiotics ranges from 0.11 to 0.13% (Kermanshahi et al., 2017). The percentage of gizzard weight found in this study ranged from 1.47%–1.67%, and is still in the normal range. According to Ramli et al. (2008), the percentage of gizzard weight ranges from 1.88%-2.23% of live weight whereas Kokoszynski et al. (2017) reported that broiler gizzard weight ranges from 1.20%-1.30%. The gizzard weight obtained, however, is lower than that reported by Malik et al. (2016) in broiler chickens fed with 0.05% probiotic, 0.20% acidifier, and their combination, namely 2.37% to 2.56%. Ali et al. (2018) also reported higher gizzard weight (2.06%) in broilers fed with organic acids as a substitute for antibiotics.

The average percentage of broiler liver weight in this study ranged from 2.24%-2.54%. According to Khotimah (2002), the normal percentage of liver weight ranges from 2.15%-2.59%. Another study reported that the average percentage of broiler liver is 2.61-2.78% of the live weight whereas the results of study performed by Natsir (2008) showed that the relative liver weight of broiler chicken fed with a combination of citric acid and lactic acid was 2.00-2.81%.

Effect of Treatment on Small Intestine Length

The effect of the addition of propionic acid in feed on the length of the small intestine of broiler chickens is presented in Table 4. The results of the analysis of variance showed that the addition of propionic acid up to 0.75% had no significant effect (P>0.05) on the length of the duodenum, jejunum, and ileum. This indicates that propionic acid does not have a negative impact on the small intestine. According to Zunita et al. (2004), the addition of organic acids in animal feed can maintain gut performance as it creates a harmonious environment for the development of beneficial microflora. A balanced microflora condition causes the formation of a good defense system in the intestinal lumen (Jepppson et al., 2004).

The average length of duodenum observed in this study ranged from 28.67-31.50 cm, while the jejunum length ranged from 70.50-77.83 cm, and the ileum ranged from 56.67-59.17 cm. The total length of the intestine in this study ranged from 155.84 to 168.50 cm. Mabelebele et al. (2014) reported that the intestinal length of the broiler chicken with Ross strain 308 was 163.10 cm (duodenum 31.90 cm, jejunum 60.60 cm, and ileum 70.60 cm). The length of the small intestine found in this study was longer compared to the results of reported by Fernandes et al. (2014) which observed that the length of small intestine of broiler chickens aged 42 days fed with several combinations of organic acids is 141.37 cm with a duodenal length 30.25 cm, jejunum 55.87 cm and ileum 55.25 cm. Meanwhile, the results of research by Adil et al. (2011) reported a longer small intestine in broiler chickens fed with several types of organic acids than those fed without organic acids supplementation. Rehman et al. (2016) reported that broilers whose feed was supplemented with acetic acid up to 0.3% had a longer small intestine (1.43%-5.44%) compared broiler chickens whose feed without acetic acid supplementation. This increase is related to the increase in the length and width of the intestinal villi (Sabour et al., 2019).

CONCLUSION

It can be concluded that the addition of propionic acid in poultry feed up to 0.75% is able to increase the percentage of live weight, but does not have a negative impact on the condition of the internal organs and the length of the small intestine of broiler chickens.

REFERENCES


